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Soil quality - Identification of ecotoxicological test species by DNA barcoding (ISO 21286:2019)

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ICS 13.080.30

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EUROPEAN STANDARD

NORME EUROPÉENNE

EUROPÄISCHE NORM

EN ISO 21286

April 2020

ICS 13.080.30

English Version

Soil quality - Identification of ecotoxicological test species by DNA barcoding (ISO 21286:2019)

Qualité du sol - Identification des espèces par codes-barres ADN dans les essais d'écotoxicologie (ISO 21286:2019)

Bodenbeschaffenheit - Allgemeine Anleitung zur Verwendung des DNA-Barcodes in ökotoxikologischen Untersuchungen (ISO 21286:2019)

This European Standard was approved by CEN on 13 April 2020.

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European foreword

The text of ISO 21286:2019 has been prepared by Technical Committee ISO/TC 190 "Soil quality" of the International Organization for Standardization (ISO) and has been taken over as EN ISO 21286:2020 by Technical Committee CEN/TC 444 "Environmental characterization of solid matrices" the secretariat of which is held by NEN.

This European Standard shall be given the status of a national standard, either by publication of an identical text or by endorsement, at the latest by October 2020, and conflicting national standards shall be withdrawn at the latest by October 2020.

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Endorsement notice

The text of ISO 21286:2019 has been approved by CEN as EN ISO 21286:2020 without any modification.

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Contents

Page

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Foreword

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The procedures used to develop this document and those intended for its further maintenance are described in the ISO/IEC Directives, Part 1. In particular the different approval criteria needed for the different types of ISO documents should be noted. This document was drafted in accordance with the editorial rules of the ISO/IEC Directives, Part 2 (see www.iso.org/directives).

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This document was prepared by Technical Committee ISO/TC 190, *Soil quality*, Subcommittee SC 4, *Biological characterization*.

Any feedback or questions on this document should be directed to the user's national standards body. A complete listing of these bodies can be found at www.iso.org/members.html.

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Introduction

Currently, test species identification is usually based on morphological characters. However, this does not always give clear results because

a) few taxonomic experts are available,

b) closely related species can differ by a few, easily overlooked characters, and

c) even more importantly, several test species are in fact complexes of cryptic species.

A good example is the compost worm *Eisenia fetida/andrei* (used in ISO 11268-1, ISO 11268-2 and ISO 17512-1), in which morphological traits alone may not be sufficient to discriminate between both species[5][36]. Another well-known case is the predatory mite, *Hypoaspis* (*Geolaelaps*) *aculeifer*[50], which might get confused with *H. miles*, widely used in biological pest control[31].

Species misidentifications, the use of a morphospecies which is actually a complex of cryptic species, or even species mixing in lab cultures, can be a serious problem for the reliability of the ecotoxicological tests. Sibling species in a morphospecies complex can exhibit ecological, behavioural, and physiological differences, and can differ also in their response to toxicants (e.g. References [2], [17], [35], [40]). This also seems to be the case of the springtail *Folsomia candida* (used in ISO 11267 and ISO 17512-2), in which considerable levels of genetic differentiation have been found among natural populations of *F. candida* and among laboratory strains[9][19][41]. Although different laboratory strains have been found to exhibit only minor differences in the sensitivity towards some chemicals[12][9], other studies have detected significant variation in phenmedipham avoidance behaviour and divergent fitness responses to cadmium exposure among genetically differentiated strains[14][30]. Moreover, even if two species have similar responses to toxicants, the presence of two species within the same laboratory culture can result in the production of sterile hybrids, which will bias the outcome of reproduction tests[36].

Implementing species identification via DNA barcoding can help to overcome these obstacles, ensuring that the species or strain used for testing is well characterized. As a result, quality assurance can be improved, making the results obtained by different ecotoxicological laboratories far more reliable and comparable. For *Eisenia fetida/E. andrei* this work, including an international ringtest, has already been performed[36], see Annex A. The conclusions of this ringtest can be summarized as follows.

— DNA barcoding is a reliable and practical method for identifying *Eisenia* species.

— Only 17 out of 28 ecotoxicological laboratories were correct in their taxonomic assignment. Most laboratories with wrong or unknown assignments actually have *E. andrei* in stock.

— The existence of a cryptic species pair within *E. fetida* is a plausible hypothesis.

— It is important that earthworms used for ecotoxicological tests are regularly (re-)identified by DNA barcoding.

Very probably, similar experiences and recommendations can be drawn for other invertebrates species used in terrestrial ecotoxicology, as well as plants. Indeed, DNA barcoding has proven to be useful for specimen identification and species delimitation in many organism groups, including other earthworms[13][37], enchytraeids[16], mites[15], collembolans[32], molluscs[42], nematodes[28] and terrestrial plants[8].

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1 Scope

This document specifies a protocol to identify ecotoxicological test specimens (mainly invertebrates and plants) to the species level, based on the DNA barcoding technique. This protocol can be used by laboratories performing DNA barcoding in order to standardize both the wet-lab and data analysis workflows as much as possible, and make them compliant with community standards and guidelines.

This document does not intend to specify one particular strain for each test method, but to accurately document the species/strain which was used.

NOTE 1 This does not imply that DNA barcoding is performed in parallel to each test run, but rather regularly (e.g. once a year, such as reference substance testing) and each time a new culture is started or new individuals are added to an ongoing culture.

This document does not aim at duplicating or replacing morphological-based species identifications. On the contrary, DNA barcoding is proposed as a complementary identification tool where morphology is inconclusive, or to diagnose cryptic species, in order to ensure that the results obtained from different ecotoxicological laboratories are referring to the same species or strain.

This document is applicable to identifications of immature forms which lack morphological diagnostic characters (eggs, larvae, juveniles), as well as the streamline identification of specimens collected in field monitoring studies, where large numbers of organisms from diverse taxa are classified.

NOTE 2 In principle, all species regularly used in ecotoxicological testing can be analysed by DNA barcoding. Besides the earthwoms *Eisenia fetida* and *E. andrei*, further examples for terrestrial species are *Lumbricus terrestris*, *L. rubellus*, *Allolobophora chlorotica*, *Aporrectodea rosea,* and *A. caliginosa*, *Dendrodrilus rubidus, Enchytraeus albidus,* and *E. crypticus* (Haplotaxida); *Folsomia candida*, *F. fimetaria, Proisotoma minuta,* and *Sinella curviseta* (Collembola); *Hypoaspis aculeifer* and *Oppia nitens* (Acari); *Aleochara bilineata* and *Poecilus cupreus* (Coleoptera); *Scathophaga stercoraria, Musca autumnalis* (Diptera) or *Pardosa* sp. (Arachnida). Nematodes or snails and even plants can also be added to this list.

2 Normative references

There are no normative references in this document.

3 Terms and definitions

For the purposes of this document, the following terms and definitions apply.

ISO and IEC maintain terminological databases for use in standardization at the following addresses:

— ISO Online browsing platform: available at https://www.iso.org/obp

— IEC Electropedia: available at http://www.electropedia.org/

3.1
amplicon
specific DNA product generated by *PCR* (3.5) using one pair of *PCR primers* (3.6)

3.2
DNA barcode
unique pattern of DNA sequence that identifies each species

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